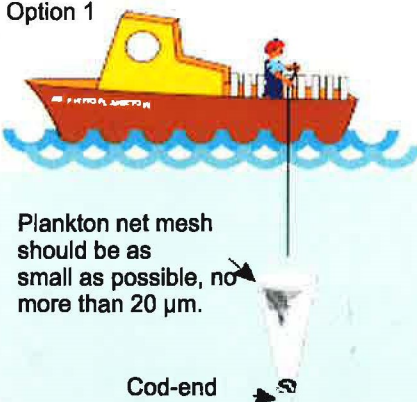


PHYTOPLANKTON SAMPLE PREPARATION

Materials Needed: 20 μm Plankton net, large bucket or hose, rope, 100 μm and 20 μm filtration, squeeze bottle, plastic dispenser, clean vials, 0.1M acetic acid

Collection: Collect phytoplankton using any one of the three options below.


1 Option 1



Plankton net mesh should be as small as possible, no more than 20 μm .

Cod-end

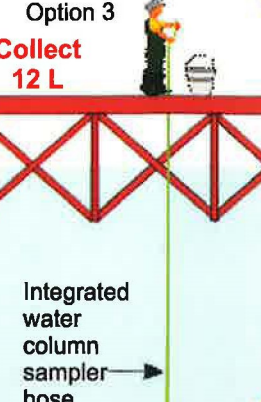
Option 2



Bucket

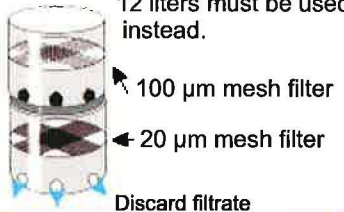
Option 3

Collect 12 L



Integrated water column sampler hose

2 Filter contents of cod-end (approximately 300 mL). When using a bucket or hose 12 liters must be used instead.

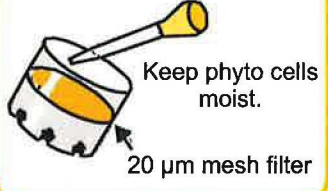


100 μm mesh filter

20 μm mesh filter

Discard filtrate


3 Hold 20 μm mesh filter at an angle and squirt gently with sea water so that cells can collect to the side.



Keep phyto cells moist.

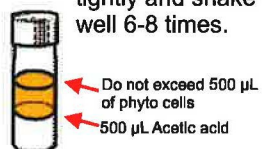
20 μm mesh filter

4 Collect the concentrated phyto cells with a plastic dispenser and transport into a clean vial.



Extraction:

1 Add phyto cells to 500 μL of 0.1 M Acetic acid so that the total volume does not exceed 1 mL. Cap tightly and shake well 6-8 times.

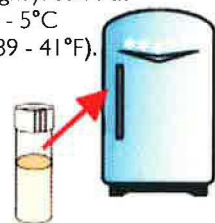


Do not exceed 500 μL of phyto cells

500 μL Acetic acid

The sample is ready to test

2 For future use keep the extract capped tightly. Store at 4 - 5°C (39 - 41°F).



Clean all equipment with 5% bleach and rinse well with tap water to avoid contaminating next sample.

Refer to Scotia Rapid Test Instruction Sheet to perform test.

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SRT08Apr14-503



ASP RAPID TEST

A qualitative lateral flow screen for the detection of ASP toxins in shellfish

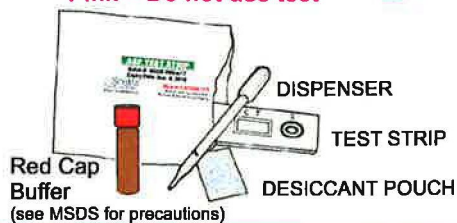
Batch 40003-01Nov16

PHYTO EXTRACTION

Field Method

Materials Needed: All materials needed are provided in the kit

- 1** •Check Expiry Date (contact if test is expired)
•Check Desiccant Pouch:
Blue = OK
Pink = Do not use test



- 2** Before using, hold buffer vial as shown, 1 inch above table.

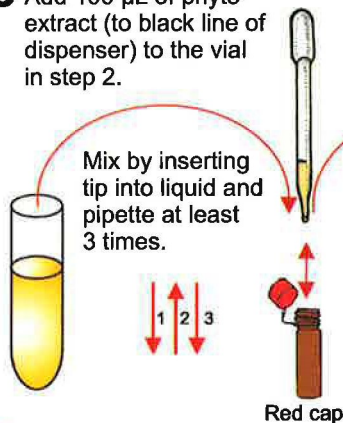


Tap 3 times on a table to get the liquid from the lid to the vial.

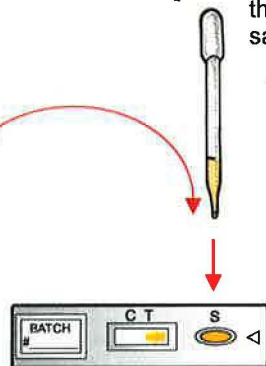
1"

Table

- 3** Add 100 μ L of phyto extract (to black line of dispenser) to the vial in step 2.



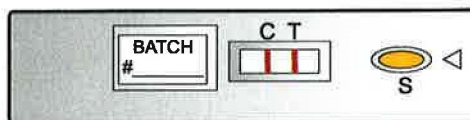
- 4** Place 100 μ L of the mixture into sample well "S".



DO NOT REUSE DISPENSERS

5

Read your results between 45-75 minutes.
Turn this page for C & T Line Interpretation.

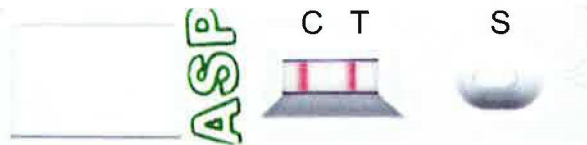


Test Interpretation

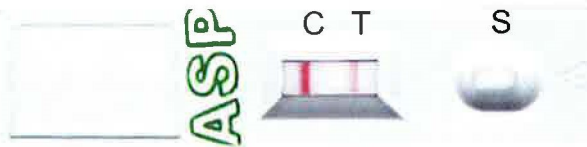
Place your cassette in the blank space indicated below.
Interpret the test result as follows:

- 1) If your T-line is darker than the Positive result below, the test is **Negative**.
- 2) If your T-line is equal to or fainter than the Positive result below, the test is **Positive**. When in doubt, consider the test **Positive**. Positive results should be confirmed with analytical testing.
- 3) If your C line is equal to or fainter than the invalid test below, your test is invalid. Repeat the assay with a new test. If the same result occurs, your sample contains an unknown agent that inhibits the tests reaction; an alternative testing method should be used.

A **Negative** Result



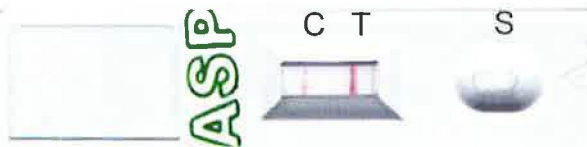
A **Positive** Result



Be Safe!

We Recommend Placing Your
Cassette Here for Comparison
Purposes

Invalid Test



Dispose of used test in the trash or keep for a permanent record.

If you need assistance, contact us at techsupp@scotiarapidtesting.ca
Technical Assistance:

- 1) Scan or photograph your problem cassettes before they completely dry.
- 2) Save and refrigerate your extract.
- 3) Call for shipping instructions before sending us extracts or tissue.

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